Microbiology

ANTIBIOTIC PROFILE OF ACINETOBACTER BAUMANNII IN TERTIARY CARE HOSPITAL, VADODARA, GUJARAT, INDIA.

Dr. Bindiya Ghedia MD Microbiology, Assistant Professor

(ABSTRACT) Acinetobacter baumannii is one of the most important bacteria in Hospital-acquired infections. Antibiotic resistance in this bacterium leads to many problems in treating patients. This prospective study was conducted on 80 isolates of Acinetobacter baumannii collected from clinical samples in the Microbiology Department, Smt. B. K. Shah Medical Institute and Research Center, Vadodara. The antibiotic resistance pattern was determined by the VITEK 2 automated machine. Out of 80 isolates, the highest rate of sensitivity was seen in Minocycline (50 %). The occurrence of infection was more common in males than females. Isolation of bacterium was more from pus samples than other samples. So the study was conducted to demonstrate the burden of micro-organisms in hospitals and their antibiotic sensitivity pattern.

KEYWORDS : Acinetobacter Baumannii, Antibiotic Sensitivity Pattern.

INTRODUCTION:

Acinetobacter baumannii is a strictly aerobic, nonmotile, Gramnegative, lactose nonfermenting, oxidase-negative, and catalasepositive bacterium. Within hospital settings, Acinetobacter baumannii has become a common pathogen, which can infect the respiratory tract, blood, soft tissues, and urinary tract of a person. It is the causative agent of nosocomial infections leading to septicemia, meningitis, endocarditis, pneumonia, wound, and urinary tract infections [1, 2]. There are 32 Acinetobacter named and unnamed species, which have been identified [3]. Acinetobacter species lead to infections, which are associated with increased morbidity and mortality rates [4, 5]. Despite the fact that the pathogen is often hospital-associated, initial infection can be transmitted by patients, admitted from other hospitals [6, 7]. About 25 years ago, A. baumannii was found to be resistant to antimicrobial agents, such as aminopenicillins, cephalosporins, first- and second-generation cephalosporins, cephamycins, aminogly cosides, ureidopenicillins, chloramphenicol, and tetracyclines. Strains of A. baumannii have started to acquire resistance against newly developed antimicrobial agents and have become prevalent in many hospitals [8]. More recently, the term "extensively drug-resistant" A. baumannii (XDRAB) is used to denote bacterial isolates resistant to all authorized antibiotics except two categories of antibiotics such as tigecycline and polymyxins [9]. The risk of A. baumannii infection from hospitalized patients is based on some important factors, such as bacterial colonization, medical staff-to-patient ratio, and other ward characteristics [10]. The current study was conducted to determine the prevalence of A. baumannii and its antibiotic profile in Dhiraj Hospital that is associated with Smt. B. K. Shah Medical Institute and Research Centre, Vadodara.

MATERIALSAND METHODS:

This prospective study was done in a tertiary care hospital in Vadodara during the period of January- June 2023. 80 non-duplicate Acinetobacter baumannii were recovered from samples such as urine, pus, sputum, wound swabs, ear swabs, blood, and endotracheal aspirate from various departments of Smt. B. K. Shah Medical Institute and Research Centre (Table 1). The clinical samples were inoculated by streak plate method on nutrient agar (Himedia, Mumbai, India), Mac Conkey agar (Himedia, Mumbai, India), and blood agar (Himedia, Mumbai, India). Blood was inoculated in the BACTEC automated machine and positive cultures were plated on blood agar, Mac Conkey agar, and nutrient agar. The isolated colonies on the different media were identified based on the morphology of the colony, Gram staining, and oxidase test, and confirmed identification was done in a VITEK 2 automated machine. VITEK 2 also gave us an antibiotic profile of the bacterium in all samples. All the media, oxidase disc, and Gram staining kits were purchased from Himedia, Mumbai, India.

RESULTS:

80 *Acinetobacter baumannii* were isolated from 221 different samples. The prevalence rate of the organism was found to be 36.20%.

 Table 1: Gender-wise distribution of Acinetobacter baumannii isolates:

Gender	No. of isolates (%)
Male	56 (70%)
Female	24 (30%)
Total	80 (100%)

Table 1 shows that among 80 Acinetobacter baumannii isolates, 56 (70%) were from males and 24 (30%) from females.

Table 2: Distribution of isolates according to type of specimen:

Type of specimen	No. of isolates (%)
Pus	26 (32.5%)
Sputum	17 (21.25%)
Urine	2 (2.5%)
ET secretion	19 (23.75%)
CSF	2 (2.5%)
Blood	12 (15%)
Ascitic fluid	1 (1.25%)
Pleural fluid	1 (1.25%)
Total	80 (100 %)

Table 2 shows that out of 80 isolates, 26 (32.5%) of the isolates were from Pus, 19(23.75%) were from ET secretion, 17(21.25%) were from sputum, 12 (15%) were from blood, 2 (2.5%) were from urine and CSF each, 1 (1.25%) were from ascitic fluid and Pleural fluid each.

Table 3: Distribution of isolates according to Department of hospital from where the sample has been received:

Department of Hospital	No. of isolates
ICU	50 (62.5%)
Surgery	10 (12.5%)
Medicine	8 (10%)
Orthopedics	3 (3.75%)
Obstetrics and Gynecology	1 (1.25%)
Casualty	5 (6.25%)
Skin	1 (1.25%)
Paediatrics	1 (1.25%)
Chemotherapy	1 (1.25%)
total	80 (100%)

Table 3 shows that 50 (62.5 %) of strains were isolated from the samples sent from ICU followed in decreasing order by 10 (12.5%) from Surgery Department, 8 (10%) from Medicine Department, 5 (6.25%) from Casualty, 3 (3.75 %) from Orthopedic Department, 1 (1.25%) from Paediatric Department, Skin, Chemotherapy, Obstetric and Gynecology Department each.

Table 4: Distribution of isolates according to their Antibiotic sensitivity pattern:

Antibiotic	Short form	No. of isolates (sensitive)	Percentage
PIPERACILLIN/TAZOBAC TAM	PIT	16	20%
CEFTAZIDIME	CAZ	11	13.75%
CEFOPERAZONE/SULBA CTAM	CFS	24	30%
CEFEPIME	CPM	12	15%
INDIAN JOURNAL OF APPLIED RESEARCH 31			

Volume - 14 Is	ssue - 05 May	- 2024	PRINT ISSN No.	2249 - 555X	DOI : 10.36106/ijar

IMIPENEM	IPM	11	13.75%
MEROPENEM	MRP	16	20%
AMIKACIN	AK	17	21.25%
GENTAMICIN	GEN	22	27.5%
CIPROFLOXACIN	CIP	11	13.75%
LEVOFLOXACIN	LE	14	17.5%
MINOCYCLIN	MI	40	50%
TIGECYCLINE	TGC	15	18.75%
COLISTIN	CL	1	1.25%
TRIMETHOPRIM/SULPHA	COT	21	26.25%
METHOXAZOLE			
Total		80	

Table 4 shows that out of 80 *Acinetobacter baumannii* isolates, 40 isolates are most sensitive to Minocyclin (50%), followed in decreasing order by Cefoperazone/Sulbactem (30%), Gentamicin (27.5%), Trimethoprim/Sulphamethoxazole (26.25%), Amikacin (21.25%), Meropenem (20%), Pipercillin/Tazobactam (20%), Tigecycline (18.75%), Levofloxacin (17.5%), Cefepime (15%), Ceftazidime (13.75%), Imipenem (13.75%), Ciprofloxacin (13.75%), Colistin (1.25%), Aztreonam and Fosfomycin (0%). 92.5 % isolates were found to be multidrug resistant.

DISCUSSION:

Acinetobacter baumannii is one of the most common cause of hospitalacquired infections among Gram-negative bacteria. Widespread use of antimicrobials has resulted in the emergence of multidrug-resistant isolates among these micro-organisms. Multiple drug resistance (MDR) is the resistance exhibited by a microorganism to at alteast one antibiotic in three or more antibiotic categories. In our study, out of 80 isolates, 74 (92.5 %) isolates are MDR. The high proportion of multidrug-resistant A. baumannii has been reported in other studies globally with major impact reported in Asian countries including Malaysia, India, and Pakistan [11,12,13]. It causes infections frequently in clinical settings, especially in surgical wards and ICUs, and the resistance patterns in different geographical areas. Hence antibiotic surveillance is of prime importance to the policy makers to frame the empirical treatment strategy for these bacterial infections. The prevalence of isolation was higher from ICU and surgery Department samples which is comparable to studies done by Santosh Kumar Yadav et al [14] and George et al [15]. Acinetobacter baumannii occurrence was predominant in males (70 %) in our study which is higher than the study done by Santosh Kumar et al [14]. In this study, the frequency of Acinetobacter baumannii was predominant in pus (32.5 %) than in other specimens which was similar to studies of Aroma et al and Mishra et al [16,17]. In the present study, the Prevalence rate of Acinetobacter baumannii was 36.20 % which is comparable to the study done by Anthony et al [18]. In our study, the isolates are most sensitive to Minocycline (50 %). Recently use of Minocycline for the treatment of Acinetobacter Baumanii has increased due to its high susceptibility [19].

CONCLUSION:

Acinetobacter baumannii is one of the most common nosocomial pathogens and extensive use of antimicrobial agents has led to the emergence of multidrug-resistant isolates in hospital settings particulary its incidence is high in ICU patients and patients of surgery Department. This may be due to prolonged stay in hospital. Infection with Acinetobacter Baumannii is more common in males than females. Minocycline is one of the most active agent for the treatment of Acinetobacter infections.

REFERENCES:

- Muray C. K. and Hospenthal D. R., (2005). "Treatment of multidrug resistant *Acinetobacter*," *Current Opinion in Infectious Diseases, vol. 18*, no. 6, pp. 502–506,
 Joly-Guillou M. L., (2005). "Clinical impact and pathogenicity of *Acinetobacter*,"
- Joy-Guinou M. L., (2005). Clinical impact and panogenicity of Acinetobacter, Clinical Microbiology and Infection, vol. 11, pp. 868–873,.
 Dijkshoom L., Nemec A., and Seifert H., (2007). "An increasing threat in hospitals:
- [3] Dijkshoorn L., Nemec A., and Seifert H., (2007). "An increasing threat in hospitals: multidrug-resistant Acinetobacter baumannii," Nature Reviews Microbiology, vol. 5, no. 12, pp. 939–951.
 [4] Kramer A., Schwebke I., and Kampf G., (2006). "How long do nosocomial pathogens
- [4] Kramer A., Schwebke I., and Kampf G., (2006). "How long do nosocomial pathogens persist on innanimate surfaces? A systematic review," *BMC Infectious Diseases, vol. 6*, no. 1, p. 130,.
- [5] Young L. S., Sabel A. L., and Price C. S., (2007). "Epidemiologic, clinical, and economic evaluation of an outbreak of clonal multidrug-resistant Acinetobacter baumannii infection in a surgical intensive care unit," *Infection Control & Hospital Epidemiology*, vol. 28, no. 11, pp. 1247–1254,
- vol. 28, no. 11, pp. 1247–1254,.
 [6] Montefour K., Frieden J., Hurst S., (2008). "Acinetobacter baumannii: an emerging multidrug-resistant pathogen in critical care," Critical Care Nurse, vol. 28, no. 1, pp. 15–25.
- Bernards A. T., Frenay H. M., Lim B. T., (1998). "Methicillin resistant Staphylococcus aureus and Acinetobacter baumannii: an unexpected difference in epidemiologic
 - 32 INDIAN JOURNAL OF APPLIED RESEARCH

- behavior," American Journal of Infection Control, vol. 26, no. 6, pp. 544–551.
 [8] Al Jarousha A. M., El Jadba A. H., Al Afifi A. S., and El Qouqa I. A., (2009). "Nosocomial multidrug-resistant Acinetobacter baumannii in the neonatal intensive care unit in Gaza City, Palestine," International Journal of Infectious Diseases, vol. 13, no. 5, pp. 623–628.
- [9] Çelik C., Gozel M., Dayı F., and G'ult'urk E. , (2014), "Increasing " antimicrobial resistance in nosocomial pathogens; multidrugresistant extensively drug-resistant and pandrug-resistant Acinetobacter baumannii," Journal of Microbiology and Infectious Diseases, vol. 4, no. 1, pp. 7–12.
- [10] Abbo A., Navon-Venezia S., Hammer-Muntz O., Krichali T., Siegman-Igra Y., and Carmeli Y., (2005). "Multidrug-resistant Acinetobacter baumannii," Emerging Infectious Diseases, vol. 11, no. 1, pp. 22–29.
- [11]. Dash M, Padhi S, Pattnaik S, Mohanty I, Misra P. (2013). Frequency, risk factors and antibiogram of *Acinetobacter* species isolated from various clinical samples in a tertiary care hospital in Odisha, India. *Avicenna J Med. Oct*; 3(4):97–102. [PMC free article] [PubMed] [Google Scholar]
- [12] Nazmul MHM, Jamal H, Fazlul MKK. (2012). Acinetobacter species-associated infections and their antibiotic susceptibility profiles in Malaysia. Biomed Res. 23:571–5. [Google Scholar]
- Biglari S, Hanafah A, Ramli R, Rahman MM, Nizam Khaithir TM. (2013). Clinicoepidemiological nature and antibiotic susceptibility profile of *Acimetobacter* species. *Pakistan J Med Sci. Apr;29*(2):469–73. [PMC free article] [PubMed] [Google Scholar].
 Yadav S. K., ^{1,2} Bhujel R., ² Hamal P.,³ Mishra S.K., ²Sharma S.,² and Sherchand J. B.³
- [14]. Yadav S. K., ¹² Bhujel R., ² Hamal P., ³ Mishra S.K., ² Sharma S., ² and Sherchand J. B.² (2020). Burden of Multidrug-Resistant Acinetobacter baumannii Infection in Hospitalized Patients in a Tertiary Care Hospital of Nepal. Infect Drug Resist. ; 13: 725-732.
- [15]. Eliopoulos G. M., Lisa L. Maragakis, Trish M. P. (2008). Acinetobacter baumannii: Epidemiology, Antimicrobial Resistance, and Treatment Options. Clinical Infectious Diseases, Volume 46, Issue 8, 15 April, Pages 1254–1263, https://doi.org/10. 1086/529198.
- [16]. Oberoi A., Aggarwal A., Lal M. (2009). A Decade of an Underestimated Nosocomal
- Pathogen-Acinetobacter In a Tertiary Care Hospital in Punjab. Vol. 11 No. 1.
 [17]. Mishra B, Bhujwala RA, Shriniwas (1986). Nonfermenters in human infections. Ind J Med Res 83: 561-566.
- [18]. Anthony D. Harris, MD, MPH'; Lisa Pineles, MA'; J. Kristie Johnson, PhD'. (2023). Prevalence of Acinetobacter baumannii and Candida auris in Patients Receiving Mechanical Ventilation. JAMA. 330(18):1769-1772. doi:10.1001/jama.21083.
- [19]. Tarazi Z., Sabet M., Dudley M. N., Griffith D. C.. Pharmacodynamics of Minocycline against *Acinetobacter baumannii* in a Rat Pneumonia Model. DOI: https://doi.org/10. 1128/aac.01671-18.